

Notification Date: February 22, 2022 Effective Date: March 15, 2022

Friedreich Ataxia, Repeat Expansion Analysis, Varies

Test ID: AFXN

Useful for:

Molecular confirmation of clinically suspected Friedreich ataxia

Reflex Tests:

Test ID	Reporting Name	Available Separately	Always Performed
CULFB	Fibroblast Culture for Genetic Test	Yes	No
CULAF	Amniotic Fluid Culture/Genetic Test	Yes	No
MATCC	Maternal Cell Contamination, B	Yes	No

Methods:

Polymerase Chain Reaction (PCR)

Reference Values:

FXN

Normal alleles: <34 GAA repeats Borderline alleles: 34-65 GAA repeats Expanded alleles: >65 GAA repeats

An interpretive report will be provided.

Specimen Requirements:

Patient Preparation: A previous bone marrow transplant from an allogenic donor will interfere with testing. Call 800-533-1710 for instructions for testing patients who have received a bone marrow transplant.

Submit only 1 of the following specimens:

Specimen Type Whole Blood

Container/Tube:

Preferred: Lavender Top (EDTA) or yellow top (ACD)

Acceptable: Any anticoagulant

Specimen Volume: 3 mL

Collection Instructions: 1. Invert several times to mix blood.

2. Send whole blood specimen in original tube. Do not aliquot.

Specimen Stability Information: Ambient (preferred)/Refrigerated

Minimum Volume: 0.5 mL

Specimen Type Amniotic fluid

Container/Tube: Amniotic fluid container

Specimen Volume: 20 mL

Specimen Stability Information: Ambient (preferred)/Refrigerated

Minimum Volume: 10 mL

Specimen Type Chorionic villi

Container/Tube: 15-mL tube containing 15 mL of transport media

Specimen Volume: 20 mg

Specimen Stability Information: Refrigerated

Minimum Volume: 5 mg

Specimen Type Confluent cultured cells

Container/Tube: T-25 flask
Specimen Volume: 2 Flasks

Collection InstructionsSubmit confluent cultured cells from another laboratory

Specimen Stability Information: Ambient (preferred)/Refrigerated

Specimen Type Blood spot

Supplies: Card – Blood Spot Collection (Filter Paper) (T493)

Container/Tube:

Preferred: Collection card (Whatman Protein Saver 903 Paper)

Acceptable: Perkin/Elmer 266 (formerly Ahlstrom 226) filter paper, or Blood Spot

Collection Card

Specimen Volume: 5 Blood spots

Collection Instructions: 1. An alternative blood collection option for a patient older than 1 year of

age is a fingerstick. See Dried Blood Spot Collection Tutorial for how to

collect blood spots via fingerstick: https://vimeo.com/508490782.

2. Let blood dry on the filter paper at ambient temperature in a horizontal

position for a minimum of 3 hours.

3. Do not expose specimen to heat or direct sunlight.

4. Do not stack wet specimens.

5. Keep specimen dry.

Specimen Stability Information: Ambient (preferred)/Refrigerated

Minimum Volume: 5 punches, 3-mm diameter

Additional Information: 1. For collection instructions, see <u>Blood Spot Collection Instructions</u>.

2. For collection instructions in Spanish, see Blood Spot Collection Card-

Spanish Instructions (T777).

3. For collection instructions in Chinese, see Blood Spot Collection Card-

Chinese Instructions (T800).

4. Due to lower concentration of DNA yielded from blood spots, it is possible that additional specimen may be required to complete testing.

Note:

Due to its complexity, prenatal testing will not be accepted without approval from the laboratory. Call 800-533-1710 for prenatal testing consultation. Prenatal specimens can be sent Monday through Thursday and must be received by 5 p.m. CST on Friday in order to be processed appropriately. All prenatal specimens must be accompanied by a maternal blood specimen. Order MATCC / Maternal Cell Contamination, Molecular Analysis, Varies on the maternal specimen.

Specimen Stability Information:

Specimen Type		Temperature	Time	Special Container
	Varies	Varies		

Cautions:

For familial testing, it is important to first document the molecular etiology of disease in an affected family member to confirm that a repeat expansion is the underlying mechanism of disease in the family. Specifically, this assay will not detect nonrepeat expansion variants (eg, sequence variants, deletions, and duplications).

It is strongly recommended that patients undergoing genetic testing receive genetic counseling.

Test results should be interpreted in the context of clinical findings, family history, and other laboratory data, such as frataxin concentrations (see FFRWB / Friedreich Ataxia, Frataxin, Quantitative, Blood and FFRBS / Friedreich Ataxia, Frataxin, Quantitative, Blood Spot). Errors in test interpretation may occur if the provided information is inaccurate or incomplete.

Rare variants (ie, polymorphisms) may exist, such as intron 1 deletions, which could lead to false-negative results. If GAA-repeat expansion results do not match clinical findings, additional testing should be considered.

Due to somatic mosaicism, GAA repeat-sizes in peripheral blood specimens may not reflect GAA repeat-sizes in other tissues (eg, central nervous system).

Bone marrow transplants from allogenic donors will interfere with testing. Call Mayo Clinic Laboratories at 800-533-1710 for instructions for testing patients who have received a bone marrow transplant.

CPT Code:

81284

81265-Maternal Cell Contamination (if appropriate)

88233- Fibroblast Culture (if appropriate) 88235-Amniotic Fluid Culture (if appropriate) 88240-Cryopreservation (if appropriate)

Day(s) Performed: Monday & Wednesday Report Available: 8 to 16 days

Note:

The following referral test code(s) will become obsolete.

Test Name	Test ID	Referral Lab Code	Referral Lab
Friedreich Ataxia Repeat Expansion Analysis - Unknown Mutation	FFRED	6031	Baylor Medical Genetics Laboratories

Questions